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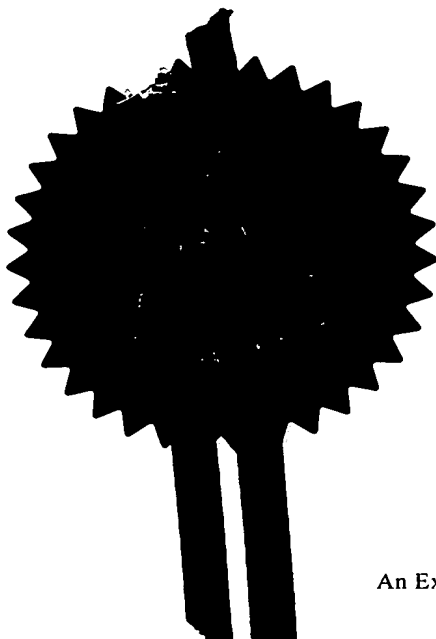
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Request for grant of a patent



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Cardiff Road
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1. Your reference TSJ/IF/32602

2. Patent application number **9826143.1** **27 NOV 1998**

3. Full name, address and post code of the or each applicant
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Patents ADP number

If the applicant is a corporate body, give the country/state of its incorporation

7557887001.

4. Title of the invention TUMOUR REJECTION ANTIGENS

5. Name of your agent VENNER, SHIPLEY & CO

"Address for service" in the United Kingdom to which all correspondence should be sent
20 LITTLE BRITAIN
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Patents ADP 1669004

6.	If you are declaring priority from one or more earlier patent applications, give the country and the date of filing of the or each of these earlier applications and the or each application number	Country	Priority application number	Date of filing

7.	If this application is divided or otherwise derived from an earlier UK application, give the number and filing date of the earlier application	Number of earlier application	Date of Filing

8. Is a statement of inventorship and of right to grant of a patent required in support of this request? YES

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11. I/We request the grant of a patent on the basis of this application.

Signature
Timothy Jump

Date
27 November 1998

12. Name and daytime telephone number of person to contact in the United Kingdom
TIMOTHY JUMP
0171-600 4212

Tumour rejection antigens

Description

This invention relates to polypeptides and proteins expressed in tumour cells and to
5 nucleic acid molecules coding for such polypeptides and proteins. The invention
also relates to expression vectors and host cells for expressing such polypeptides
and proteins, and to polypeptide-binding agents which selectively bind or are
specific for such polypeptides or proteins. The invention further relates to methods
of treating and diagnosing disease, preferably cancers, using such polypeptides,
10 proteins, nucleic acids, polypeptide-binding agents, expression vectors or
transformed host cells.

The phenotypic changes which distinguish a tumour cell from its normal
counterpart are often the result of one or more changes to the genome of the cell.
15 The genes which are expressed in tumour cells, but not in normal counterparts, can
be termed "tumour specific" or "tumour associated" genes. These tumour specific
or associated genes can be markers for the tumour phenotype.

The process by which the mammalian immune system recognises and reacts to
20 foreign or alien materials is a complex one. An important facet of the system is the
response of cytolytic T lymphocytes (CTLs) or T cells. CTLs recognise and interact
with complexes of cell surface molecules, referred to as human leukocyte antigens
("HLA"), or major histocompatibility complex molecules ("MHC" molecules), and
other peptides derived from larger molecules from within the cells carrying the
25 HLA/MHC complexes. See in this regard Male et al., Advanced Immunology (J.P.
Lipincott Company, 1987), especially chapters 6-10 and C.A. Janeway et al. Immuno
Biology third ed. (Current Biology Ltd. 1997). The interaction of T cells and
complexes of HLA/peptide is restricted, requiring a T cell specific for a particular
combination of an HLA molecule and a peptide. If a specific CTL is not present,
30 there is no T cell response even if its partner complex is present. Similarly, there is
no response if the specific complex is absent, but the CTL is present. The
mechanism is involved in the immune system's response to foreign materials, in
autoimmune pathologies, and in responses to cellular abnormalities. Much work has

focused on the mechanisms by which proteins are processed into the HLA binding peptides. See, in this regard, Barinaga, *Science* 257: 880, 1992; Fremont et al., *Science* 257: 919, 1992; Matsumura et al., *Science* 257: 927, 1992; Latron et al., *Science* 257: 964, 1992.

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The mechanism by which T cells recognise cellular abnormalities has also been implicated in cancer. A number of families of genes which are processed into peptides that are presented as HLA/peptide complexes on the surface of tumour cells, with the result that the cells can be lysed by specific CTLs, have been
10 discovered. These genes are said to code for "tumour rejection antigen precursors" or "TRAP" molecules, and the peptides derived therefrom that complex with HLA are referred to as "tumour rejection antigens" or "TRAs". Intensive efforts have been made in this field and a wealth of human tumour rejection antigens (both TRAPs and TRAs) which are recognised by T cells have been identified (Van den
15 Eynde, B.J., and P. van der Bruggen, 1997, *Curr. Opin. Immunol.* 9:684.). Among them, a TRAP encoded by the gene *MAGE-1* was initially defined by cultivating blood lymphocytes of patient MZ2 in the presence of a melanoma cell line derived from the same patient. A panel of CTL clones was generated by mixed lymphocyte-tumour cell culture (MLTC) techniques, and one of these clones recognised a
20 nonapeptide TRA derived from the *MAGE-1* TRAP, which is presented by HLA-A1 (van der Bruggen, P., C. et al., 1991, *Science (Wash. DC)*. 254:1643-1647; Traversari, C., et al., 1992, *J. Exp. Med.* 176:1453-1457 and WO92/20356). It was found later that *MAGE-1* belongs to a family of at least seventeen related genes, namely *MAGE-1* to -12 (now named *MAGE-A1* to -A12) (De Plaen, E., et al., 1994,
25 *Immunogenetics*. 40:360-369.), *MAGE-B1* to -B4 (Muscатели, F., et al., 1995, *Proc. Natl. Acad. Sci. USA*. 92:4987-4991; Dabovic, B., et al., 1995, *Mammalian Genome*. 6:571-580; and Lurquin, C., et al., 1997, *Genomics*. 46:397-408), and *MAGE-C1* (Lucas, S., et al., 1998, *Cancer Res.* 58:743-752).

30 Genes of this family are expressed in various tumours of different histological types, but are completely silent in normal tissues with the exception of testis and placenta (De Plaen, E., et al., 1994, *Immunogenetics*. 40:360-369; Dabovic, B., et al., 1995, *Mammalian Genome*. 6:571-580; Lurquin, C., et al., 1997, *Genomics*. 46:397-408; and

Lucas, S., et al., 1998, *Cancer Res.* 58:743-752.). However, as testicular germ cells and placental trophoblasts do not express MHC class 1 molecules (Haas, G.G.Jr., et al., 1988, *Am. J. Reprod. Immunol. Microbiol.* 18:47-51.), gene expression in these tissues should not lead to antigen expression. Indeed, immunisation of male mice
5 with an antigen encoded by mouse P1A gene, which has the same expression pattern as human *MAGE* gene, i.e., expressed in tumours, testis and placenta, but silent in other normal tissues, produced strong P1A-specific CTL responses that did not cause testis inflammation or alteration of fertility (Uyttenhove, C., C. et al., 1997, *Int. J. Cancer.* 70:349-356.). Antigens encoded by *MAGE* genes are, therefore,
10 suitable candidates for vaccine-based immunotherapy of cancers and as markers for providing a means of identifying a cell as a so treatable tumour cell.

So far, however, it has only proven possible to identify TRAs encoded by *MAGE-A1*, *-A3* and *-A6* by using autologous CTLs derived from mixed lymphocyte-tumour cell cultures (MLTC) and previous gene expression assays have suggested
15 that *MAGE-A10* was expressed in tumours at a level that was too low to be sufficient for CTL recognition. All these CTLs were generated from only one patient, MZ2 (Traversari, C., et al., 1992, *J. Exp. Med.* 176:1453-1457; van der Bruggen, P., et al., 1994, *Eur. J. Immunol.* 24:2134-2140; Gaugler, B., et al., 1994,
20 *Exp. Med.* 179:921-930; De Plaen, E., et al., 1994, *Immunogenetics.* 40:360-369; and P. van der Bruggen, unpublished data). However, the inventors have now been able to obtain autologous CTL clones from another melanoma patient, LB 1751, which recognize and have allowed the identification of hitherto unknown HLA-A2.1-presented TRAs encoded by *MAGE-A10* and *MAGE-A8*.

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Accordingly, the present invention provides a polypeptide comprising an unbroken sequence of amino acids from SEQ. ID. NO. 1 or 2 which has an ability to complex with an MHC molecule type HLA-A2, preferably HLA-A2.1. Polypeptides in accordance with the invention can comprise unbroken sequences of amino acids
30 from SEQ. ID. NO. 1 or 2 which have an ability to elicit an immune response from human lymphocytes.

Polypeptides in accordance with the invention can comprise nonapeptides having an unbroken sequence of amino acids from SEQ. ID. NO.1, or 2, wherein the amino acid adjacent to the N-terminal amino acid is L or M, preferably L, and the C-terminal amino acid is L, V or I, preferably L. Alternatively, the inventive polypeptide can be a decapeptide comprising a nonapeptide as defined above and, preferably an unbroken sequence of amino acids from SEQ. ID. NO. 1, or 2. In preferred embodiments the nonapeptide has the amino acid sequence GLYDGMEHL or GLYDGREHS, preferably GLYDGMEHL. In embodiments, the decapeptide can have the amino acid sequence GLYDGMEHLI or GLYDGREHSV, preferably GLYDGMPHLI.

In a further aspect, the present invention comprises a polypeptide or protein of up to about 93 amino acids in length which comprises a nonapeptide or a decapeptide as defined above. Such a polypeptide or protein can comprise an unbroken sequence of amino acids from SEQ. ID. NO.1, or 2, preferably SEQ. ID. NO. 1.

It is preferred that polypeptides in accordance with the present invention are capable of eliciting an immune response from human lymphocytes, preferably when complexed with an MHC molecule type HLA-A2, preferably HLA-A2.1. The immune response is preferably a cytolytic response from human T-lymphocytes, preferably CD8 T-cells.

In a further aspect, the present invention provides a polypeptide or protein comprising a polypeptide as defined above, wherein the amino acid sequence of said polypeptide or protein is not either of the complete sequences set out in SEQ. ID. NOs. 1 and 2, or that coded for by nucleotides 334-918 of SEQ. ID. NO. 7.

The invention also extends to polypeptides or proteins which are functionally equivalent homologues to any of the above defined polypeptides or proteins, but with the proviso that the amino acid sequence of said polypeptide or protein is not an entire sequence as set out in either of SEQ. ID. NOs. 1 and 2, or that coded for by nucleotides 334-918 of SEQ. ID. No. 7. In embodiments of the invention, the

polypeptides can be complexed with an MHC molecule type HLA-A2, preferably HLA-A2.1.

5 In another aspect, the present invention provides nucleic acid molecules, each comprising a nucleotide sequence coding for a polypeptide or protein in accordance with previously defined aspects of the invention or a complimentary nucleotide sequence, wherein said nucleotide sequence is not an entire sequence as set out in any of SEQ. ID. NOs. 3, 4, 5, 6 and 7. Such a nucleic acid molecule can comprise an unbroken sequence of nucleotides from SEQ. ID. NO. 3, 4 or 5, or a
10 complimentary sequence, or an RNA transcript of said nucleic acid molecule.

In a preferred embodiment, such a nucleic acid molecule can encode a plurality of epitopes or a polytope.

15 In a further aspect, the present invention provides expression vectors, each comprising a nucleic acid molecule as previously defined, operably linked to a promoter. Expression vectors in accordance with the invention can comprise a nucleotide sequence coding for an MHC molecule type HLA-A2, preferably HLA-A2.1, a cytokine or a co-stimulatory molecule, or a bacterial or viral genome or a
20 portion thereof.

In an additional aspect, the present invention relates to host cells, each transformed or transfected with an expression vector in accordance with the invention. Such a host cell can be transformed or transfected with an expression vector coding for an
25 MHC molecule type HLA-A2, preferably HLA-A2.1, and/or a cytokine or a co-stimulatory molecule.

In a yet further aspect, the present invention provides polypeptide-binding agents, each of which can selectively bind or is specific for an isolated polypeptide or
30 protein in accordance with the invention. A polypeptide-binding agent in accordance with the invention can comprise an antibody, preferably a monoclonal antibody or an antibody fragment specific for an isolated polypeptide in accordance with the invention. Preferably, such polypeptide-binding agents can selectively bind

or are specific for a complex of a polypeptide in accordance with the invention and an MHC molecule type HLA-A2, preferably HLA-A2.1, but do not bind said major histocompatibility molecule alone. Further polypeptide-binding agents in accordance with the invention include CTLs and CTL clones which recognise and
5 selectively lyse cells which carry a polypeptide in accordance with the invention complexed with an MHC molecule type HLA-A2, preferably HLA-A2.1.

In another aspect, the present invention relates to the use of a polypeptide or protein, isolated nucleic acid molecule, expression vector, host cell, or polypeptide-
10 binding agent in accordance with the invention, in the therapy, prophylaxis, or diagnosis of disease and, preferably, of tumours. Thus, the invention also relates to pharmaceutical compositions for the prophylaxis, therapy or diagnosis of disease, preferably of tumours, comprising a polypeptide or protein, a nucleic acid molecule, an expression vector, a host cell, or a polypeptide-binding agent in accordance with
15 the invention, optionally in admixture with a pharmaceutically acceptable carrier and optionally further comprising a major histocompatibility molecule type HLA-A2, preferably HLA-A2.1. Such pharmaceutical compositions can be employed as anti-tumour vaccines. Optionally pharmaceutical compositions in accordance with the invention can include other TRAs or TRAPs, expression vectors or host cells
20 expressing other TRAs or TRAPs, or polypeptide-binding agents specific for other TRAs or TRAPs. In another embodiment, pharmaceutical compositions in accordance with the invention can further comprise a co-stimulatory molecule.

In a preferred embodiment, a pharmaceutical composition in accordance with the
25 invention comprises an antigen presenting cell (APC), preferably a dendritic cell, which has been pulsed with a polypeptide in accordance with the invention so as to present on its surface said peptide as a complex with a major histocompatibility molecule, HLA.

30 In another aspect, the present invention provides peptide-pulsed antigen presenting cells.

In a yet further aspect, the invention relates to a method of diagnosing disease, preferably cancer, comprising contacting a biological sample isolated from a subject with an agent that is specific for a polypeptide or protein in accordance with the invention, or a nucleic acid molecule in accordance with the invention and assaying
5 for interaction between the agent and any of the polypeptide, protein or nucleic acid molecule in the sample as a determination of the disease. The polypeptide-binding agent employed in this aspect of the invention can be a polypeptide-binding agent in accordance with a previously described aspect of the invention.

10 The invention also relates to methods of producing cytolytic T-cell cultures reactive against tumour cells. Such a method can comprise steps of removing a lymphocyte sample from an individual and then culturing the lymphocyte sample with a polypeptide or protein in accordance with the invention, an expression vector in accordance with the invention, or a host cell in accordance with the invention.
15 Products comprising cytolytic T-cells reactive against a tumour cell expressing an antigen comprising a polypeptide or protein in accordance with the invention, can be used in the prophylaxis, therapy or diagnosis of disease preferably of tumours, are also encompassed in the present invention, particularly when obtained or obtainable by the aforementioned method.

20 As set out above, the present invention can involve the use of expression vectors to transform or transfect host cells and cell lines. Thus, a coding DNA sequence in accordance with the invention can be introduced into an expression vector suitable for directing expression of a polypeptide or protein in accordance with the
25 invention (coded for by that DNA sequence) in a host cell. Suitable vectors include bacterial plasmids, phage DNA, cosmids, yeast plasmids and viral DNA, such as pox virus (e.g. vaccinia), retrovirus, baculovirus and adenovirus DNA. The procedure generally involves inserting a DNA sequence to be expressed into an appropriate restriction endonuclease site so that it is operably linked to a promoter for directing
30 mRNA synthesis. A coding sequence and regulatory sequence, such as a promoter sequence, are considered to be "operably" linked when they are covalantly linked in such a way as to place the expression or transcription of the coding sequence under the influence or control of the regulatory sequence. The resulting vector may then

be employed to transform or transfect an appropriate host cell to cause that host cell to express the required polypeptide or protein. The appropriate host cells can be higher eukaryotic cells, such as mammalian cells and insect cells or can be lower eukaryotic cells, such as yeast cells, or prokaryotic cells, such as bacterial cells. Examples include E-coli, Bowes melanoma, CHO and COS cells. Selection of an appropriate host and the manner in which the vector is introduced into the host cell are matters within the knowledge of those skilled in the art. However appropriate techniques, cloning and expression vectors for use with prokaryotic and eukaryotic hosts are described in Sambrook et al, Molecular Cloning, A Laboratory Manual, Second edition, Coldspring Harbour, NY 1989.

Expression vectors in accordance with the invention can include a nucleic acid sequence coding for the HLA molecule that presents a particular polypeptide in accordance with the invention. Alternatively, the nucleic acid sequence coding for the HLA molecule can be contained within a separate expression vector within a host cell in accordance with the invention. In a situation where the vector contains both coding sequences, the single vector can be used to transfect the cell which does not normally express either one. Where the coding sequence for the inventive polypeptide or protein and the HLA molecule which presents the former are contained on separate expression vectors, the expression vectors can be cotransfected. Sequences coding for polypeptides or proteins in accordance with the invention may be used alone, when, e.g. the host cell already expresses an HLA molecule which presents the TRA.

Preferred systems for mRNA expression in mammalian cells include the pRc/CMV (available from Invitrogen, Carlsbad CA) system that contains a selectable marker such as a gene that confers G418 resistance (which facilitates the selection of stably transfected cell lines) and the human cytomegalovirus (CMV) enhancer-promoter sequences. Additionally, suitable for expression in primate or canine cells lines is the pCEP4 vector (Invitrogen), which contains an Epstein Barr virus (EBV) origin of replication, facilitating the maintenance of plasmid as a multicopy extrachromosomal element. Another expression vector is the pEF-BOS plasmid containing the promoter of polypeptide Elongation Factor 1 α , which stimulates

efficiently transcription *in vitro*. The plasmid is described by Mishizuma and Nagata (Nuc. Acids Res. 18: 5322, 1990), and its use in transfection experiments is disclosed by, for example, Demoulin (Mol. Cell. Biol. 16: 4710-4716, 1996). A further preferred expression vector is an adenovirus, described by Stratford-Perricaudet, which is defective for E1 and E3 proteins (J. Clin. Invest. 90:626-630, 1992). The use of the adenovirus as an adeno-P1A recombinant is disclosed by Warnier et al: in Intradermal injection in mice for immunisation against P1A (Int. J. Cancer, 67:303-310, 1996).

10 As stated above, the invention can involve polypeptide-binding agents specific for or selective for polypeptides or proteins in accordance with the invention. An agent should be considered as "specific" for a particular polypeptide or protein if it is capable of interacting with that polypeptide or protein in a manner which can be distinguished from its interaction with other molecules in the context in which it is
15 used. For example, such an agent may be capable of selectively binding to a relevant polypeptide or protein under the conditions prevalent in a particular assay. The term "contacting" means that a biological sample is placed in sufficient proximity to an agent and under appropriate conditions of, for example, concentration, temperature, time, to allow the specific interaction between the agent
20 and any polypeptide or protein for which it is specific, to take place. Appropriate conditions for contacting agents and biological samples are well known to those skilled in the art and are selected to facilitate the specific interaction between particular target molecules and specific agents. Polypeptide-binding agents can be used in this way in screening assays to detect the presence or absence of proteins or
25 polypeptides in accordance with the invention and in purification protocols to isolate such proteins and polypeptides. Polypeptide-binding agents in accordance with the invention can be in the form of immobilised antibodies attached to a substrate and the inventive method of diagnosing disease can involve a conventional enzyme-linked immunosorbent assay (ELISA) carried out on a protein containing
30 biological sample derived from a patient. Alternatively, the method can comprise a Western blot in which the agent is a labelled antibody and the biological sample comprises proteins derived from a patient and separated by electrophoresis on an SDS polyacrylamide gel. Polypeptide-binding agents can be used to selectively

target drugs, toxins or other molecules to cancer cells which present polypeptides in accordance with the invention. In this manner, cells present in tumours which express polypeptides or proteins in accordance with the invention can be treated with cytotoxic compounds.

5

As stated, the invention can involve antibodies or fragment of antibodies having the ability to selectively bind to polypeptides or proteins in accordance with the invention. Such antibodies include polyclonal and monoclonal antibodies, prepared according to the conventional methodology.

10

The antibodies of the present invention can be prepared by any of a variety of methods, including administering protein, fragments of protein, cells expressing the protein or fragments thereof and the like to an animal to induce polyclonal antibodies. The production of monoclonal antibodies is according to techniques well known in the art. Such antibodies may be used for example to identify tissues expressing protein or to purify protein. Antibodies also may be coupled to specific labelling agents for imaging or to antitumour agents, including, but not limited to, methotrexate, radioiodinated compounds, toxins such as ricin, other cystostatic or cytolytic drugs, and so forth. Antibodies prepared according to the invention also preferably are specific for the TRA/HLA complexes described herein.

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Significantly, as is well known in the art, only a small portion of an antibody molecule, the paratope, is involved in the binding of the antibody to its epitope (see, in general, Clark W.R. (1986) The Experimental Foundations of Modern Immunology Wiley & Sons, Inc., New York; Roitt, I. (1991) Essential Immunology, 7th Ed., Blackwell Scientific Publications, Oxford). The pFc' and Fc regions, for example, are effectors of the complement cascade but are not involved in antigen binding. An antibody from which the pFc' region has been enzymatically cleaved, or which has been produced without the pFc' region, designated an F(ab')₂ fragment, retains both of the antigen binding sites of an intact antibody. Similarly, an antibody from which the Fc region has been enzymatically cleaved, or which has been produced without the Fc region, designated an Fab fragment, retains one of the antigen binding sites of an intact antibody molecule. Proceeding further, Fab

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fragments consist of a covalently bound antibody light chain and a portion of the antibody heavy chain denoted Fd. The Fd fragments are the major determinant of antibody specificity (a single Fd fragment may be associated with up to ten different light chains without altering antibody specificity) and Fd fragments retain epitope-binding ability in isolation.

Within the antigen-binding portion of an antibody, as is well known in the art, there are complementarity determining regions (CDRs), which directly interact with the epitope of the antigen, and framework regions (FRs), which maintain the tertiary structure of the paratope (see in general, Clark, 1986; Roitt, 1991). In both the heavy chain Fd fragment and the light chain of IgG immunoglobulins, there are four framework regions (FR1 through FR4) separated respectively by three complementarity determining regions (CDR1 through CDR3). The CDRs, and in particular the CDR3 regions, and more particularly the heavy chain CDR3, are largely responsible for antibody specificity.

It is now well established in the art that the non-CDR regions of a mammalian antibody may be replaced with similar regions of nonspecific or heterospecific antibodies while retaining the epitope specificity of the original antibody. This is most clearly manifested in the development and use of "humanised" antibodies which non-human CDRs are covalently joined to human FR and/or Fc/Fc' regions to produce a functional antibody. Thus, for example, PCT International Publication Number WO92/04381 teaches the production and use of humanised murine RSV antibodies in which at least a portion of the murine FR regions have been replaced by FR regions of human origin. Such antibodies, including fragments of intact antibodies with antigen-binding ability, are often referred to as "chimeric" antibodies.

Thus, as will be apparent to one of ordinary skill in the art, the present invention also provides for $F(ab')_2$, Fab, Fv and Fd fragments; chimeric antibodies in which the Fc and/or FR and/or CDR1 and/or CDR2 and/or light chain CDR3 regions have been replaced by homologous human or non-human sequences; chimeric $F(ab')_2$ fragment antibodies in which the FR and/or CDR1 and/or CDR2 and/or

light chain CDR3 regions have been replaced by homologous human or non-human sequences; chimeric Fab fragment antibodies in which the FR and/or CDR1 and/or CDR2 and/or light chain CDR3 regions have been replaced by homologous human or non-human sequences; and chimeric Fd fragment antibodies in which the FR and/or CDR1 and/or CDR2 regions have been replaced by homologous human or non-human sequences. The present invention also includes so-called single chain antibodies. Thus, the invention can involve polypeptides of numerous sizes and types that bind specifically or selectively to polypeptides and proteins in accordance with the invention. These polypeptides may be derived also from sources other than antibody technology. For example, such polypeptide binding agents can be provided by degenerate peptide libraries which can be readily prepared in solution, in immobilised form or as phage display libraries. Combinatorial libraries can also be synthesized of peptides containing one or more amino acids. Libraries further can be synthesized of peptiods and non-peptide synthetic moieties.

Phage display can be particularly effective in identifying binding peptides useful according to the invention. Briefly, one prepares a phage library (using e.g. m13, fd, or lambda phage), displaying inserts from 4 to about 80 amino acid residues using conventional procedures. The inserts may represent a completely degenerate or biased array. One can then select phage-bearing inserts which bind to a polypeptide or protein in accordance with the invention. This process can be repeated through several cycles of reselection of phage that bind to a polypeptide or protein in accordance with the invention. Repeated rounds lead to enrichment of phage bearing particular sequences. DNA sequence analysis can be conducted to identify the sequences of the expressed polypeptides. The minimal linear portion of the sequence that binds to a polypeptide or protein in accordance with the invention can be determined. One can repeat the procedure using a biased library containing inserts containing part or all of the minimal linear portion plus one or more additional degenerate residues upstream or downstream thereof. Thus, a polypeptide or protein in accordance with the invention can be used to screen peptide libraries, including phage display libraries, to identify and select peptide binding partners of the polypeptides of the invention. Such molecules can be used, as described, for screening assays, for diagnostic assays, for purification protocols or for targeting

drugs, toxins and/or labelling agents (e.g. radioisotopes, fluorescent molecules, etc.) to cells which express a polypeptide or protein in accordance with the invention on the cell surface. Such binding agent molecules can also be prepared to bind complexes of a polypeptide or protein in accordance with the invention and an HLA molecule by selecting the binding agent using such complexes. Drug molecules that would disable or destroy tumour cells which express such complexes are known to those skilled in the art and are commercially available. For example, the immunotoxin art provides examples of toxins which are effective when delivered to a cell by an antibody or fragment thereof. Examples of toxins include ribosome-damaging toxins derived from plant or bacterial such as ricin, abrin, saporin, Pseudomonas endotoxin, diphtheria toxin, A chain toxins, blocked ricin, etc.

The invention as described herein has a number of uses, some of which are described herein. First the invention permits the diagnosis of a disorder characterised by an expression of a polypeptide or protein in accordance with the invention. The methods can involve determining expression of the gene coding for a polypeptide or protein in accordance with the invention. In the former situation, such determinations can be carried out by any standard nucleic acid determination assay, including the polymerase chain reaction or assaying with labelled hybridisation probes, while in the latter situation, assaying with polypeptide-binding agents in accordance with the invention, such as antibodies, is preferred. An alternative method for determination is an assay for recognition of a TRA/HLA complex by a peptide-specific CTL by assaying for CTL activity. Such assays include a TNF release assay, of the type described below, a chromium release assay or a technique called ELISPOT in which CTL activity can be detected via antibody detection of IFN- γ or TNF α release (Schmittel et al (1997). J. Immunol. Methods 210:167-174 and Lalvani et al. J. Exp. Med. 126 859 (1997)).

Other TRAPs or TRAs recognised by the CTL clones described herein may be isolated by the procedures detailed herein.

A variety of methodologies well known to the skilled practitioner can be utilised to obtain isolated TRA and TRAP molecules such as those which are the subject of the present invention. The protein may be purified from cells which naturally produce the protein. Alternatively, an expression vector may be introduced into cells to
5 cause production of the protein. In another method, mRNA transcripts may be microinjected or otherwise introduced into cells to cause the production of the encoded protein. Translation of mRNA in cell-free extracts such as reticulocyte lysate system also may be used to produce protein. Peptides comprising TRAs of the invention may also be synthesised *in vitro*. Those skilled in the art can also
10 readily follow known methods for isolating proteins in order to obtain isolated TRAPs and/or TRAs derived therefrom. These include, but are not limited to, immunochromatography, HPLC, size-exclusion chromatography, ion-exchange chromatography and immune-affinity chromatography.

15 Polypeptides or proteins in accordance with the invention or complexes thereof with HLA, again in accordance with the invention, may be combined with materials such as adjuvants to produce vaccines useful in treating disorders characterised by expression of a polypeptide or protein in accordance with the invention.

20 Certain therapeutic approaches based upon the disclosure are premised on a response by the subject's immune system, leading to lysis of TRA presenting cells. One such approach is the administration of autologous CTLs specific to the complex to a subject with abnormal cells of the phenotype at issue. It is within the skill of the artisan to develop such CTLs *in vitro*. Generally, a sample of cells taken
25 from a subject, such as blood cells, are contacted with a cell presenting the complex and capable of provoking CTLs to proliferate. The target cell can be a transfectant, such as a COS cell. These transfectants present the desired complex on their surface and, when combined with a CTL of interest, stimulate its proliferation. COS cells, such as those used herein are widely available, as are other suitable host
30 cells. Specific production of a CTL is well known to one of ordinary skill in the art. One method for selecting antigen-specific CTL clones has recently been described (Altman et al., Science 274:94-96, 1996; Dunbar et al., Curr. Biol. 8:413-416, 1998), in which fluorogenic tetramers of MHC class I molecule/peptide complexes are

used to detect specific CTL clones. Briefly, soluble MHC class I molecules are folded *in vitro* in the presence of β_2 -microglobulin and a peptide antigen which binds the class I molecule. After purification, the MHC/peptide complex is purified and labelled with biotin. Tetramers are formed by mixing the biotinylated peptide-MHC complex with labelled avidin (e.g. phycoerythrin) at a molar ratio of 4:1. Tetramers are then contacted with a source of CTLs such as peripheral blood or lymph node. The tetramers bind CTLs which recognise the peptide antigen/MHC class I complex. Cells bound by the tetramers can be sorted by fluorescence activated cell sorting to isolate the reactive CTLs. The isolated CTLs then can be expanded *in vitro*. The clonally expanded autologous CTLs then can be administered to the subject. Other CTLs specific to a polypeptide or protein in accordance with the invention may be isolated and administered by similar methods.

To detail a therapeutic methodology, referred to as adoptive transfer (Greenberg, J. Immunol. 136(5): 1917, 1986; Riddell et al. Science 257:238, 1992; Lynch et al, Eur. J. Immunol. 21:1403-1410, 1991; Kast et al., Cell 59:603-614, 1989), cells presenting the desired complex are combined with peripheral blood lymphocytes containing CTLs leading to proliferation of the CTLs specific thereto. The proliferated CTLs are then administered to a subject with a cellular abnormality which is characterised by certain of the abnormal cells presenting the particular complex. The CTLs then lyse the abnormal cells, thereby achieving the desired therapeutic goal.

The foregoing therapy assumes that at least some of the subject's abnormal cells present the relevant HLA/TRA complex. This can be determined very easily, as the art is very familiar with methods for identifying cells which present a particular HLA molecule, as well as how to identify cells expressing DNA or protein of the pertinent sequences. In this case, MAGE-A10 expression could be determined, for example, by conducting a PCR assay using primers from unique parts of the MAGE-A10 DNA. Alternatively, other well known antibody based techniques can be employed to identify cells presenting a relevant TRA/HLA complex. Once cells presenting the relevant complex are identified via the foregoing screening methodology, they can be combined with a sample from a patient containing CTLs. If the complex presenting cells are lysed by the mixed CTL sample, then it can be

assumed that the TRA is being presented, and the subject is an appropriate candidate for the therapeutic approaches set forth herein.

Adoptive transfer is not the only form of therapy that is available in accordance
5 with the invention. CTLs can also be provoked *in vivo*, using a number of approaches.

One approach is the use of non-proliferative cells expressing the complex as vaccines. Such vaccines can be prepared from cells, which can be host cells in
10 accordance with the invention, that present TRA/HLA complexes on their surface. The cells used in this approach may be those that normally express the complex, such as irradiated non-proliferative tumour cells or non-proliferative transfectants etcetera. Chen et al., Proc. Natl. Acad. Sci. USA 88:110-114 (1991) exemplifies this approach, showing the use of transfected cells expressing HPV E7 peptides in a
15 therapeutic regime. Various cell types may be used. Similarly, vectors carrying one or both of the genes of interest may be used. Viral or bacterial vectors are especially preferred. For example, nucleic acids which encode a polypeptide or protein in accordance with the invention may be operably linked to promoter and enhancer sequences which direct expression of the polypeptide or protein in
20 accordance with the invention in certain tissues or cell types. The nucleic acid may be incorporated into an expression vector. Expression vectors may be unmodified extrachromosomal nucleic acids, plasmids or viral genomes constructed or modified to enable insertion of exogenous nucleic acids, such as those encoding a polypeptide or protein in accordance with the invention. Nucleic acids encoding a polypeptide
25 or protein in accordance with the invention also may be inserted into a retroviral genome, thereby facilitating integration of the nucleic acid into the genome of the target tissue or cell type. In these systems, the gene of interest is carried by a microorganism, e.g., a vaccinia virus, retrovirus or the bacteria BCG, and the materials *de facto* "infect" host cells. The cells which result present the complex of
30 interest, and are recognised by autologous CTLs, which then proliferate. In all cases where cells are used as a vaccine, these can be cells transfected with coding sequences for one or both of the components necessary to provoke a CTL response, or be cells which already express both molecules without the need for

transfection. These cells can also be antigen presenting cells (APCs), such as dendritic cells (DC) which have been "pulsed" with the TRAs of the invention or peptides derived therefrom (Nestle et al. Nat. Med. 4:328-332, 1998; Mukherji et al. Proc. Nat. Acad. Sci. USA. 92:8078-8082, 1995; Hu et al. Cancer Res. 56:2479-2483, 1996).

Vaccines also encompass naked DNA or RNA, encoding a polypeptide or protein in accordance with the invention, which may be produced *in vitro* and administered via injection, particle bombardment, nasal aspiration and other methods. Vaccines of the "naked nucleic acid" type have been demonstrated to provoke an immunological response including generation of CTLs specific for the peptide encoded by the naked nucleic acid (Science 259:1745-1748, 1993). When "disorder" is used herein, it refers to any pathological condition where the tumour rejection antigen precursor is expressed. An example of such a disorder is cancer, particularly melanoma.

A similar effect can be achieved by combining a polypeptide or protein in accordance with the invention with an adjuvant to facilitate incorporation into HLA presenting cells *in vivo*. The polypeptide or protein in accordance with the invention complexes with a molecule which presents the polypeptide or protein in accordance with the invention without the need for further processing. Generally, subjects can receive an intradermal injection of an effective amount of a polypeptide or protein in accordance with the invention. Initial doses can be followed by booster doses, following immunisation protocols standard in the art.

Especially preferred are nucleic acids encoding a series of epitopes, known as "polytopes". The epitopes can be arranged in sequential or overlapping fashion (see, e.g. Thompson et al, Proc.Natl. Acad. Sci. USA 92:5845-5849, 1995; Gilbert et al, Nature Biotechnol. 15:1280-1284, 1997) with or without the natural flanking sequences, and can be separated by unrelated linker sequences if desired. The polytope is processed to generated individual epitopes which are recognized by the immune system for generation of immune responses.

Thus, for example, peptides in accordance with the invention and which are presented by MHC molecules and recognised by CTL or T helper lymphocytes can be combined with peptides from other tumour rejection antigens (e.g. by preparation of hybrid nucleic acids or polypeptides) to form "polytopes".

5 Exemplary tumour associated peptide antigens that can be administered to induce or enhance an immune response are derived from tumour associated genes and encoded proteins including MAGE-1, MAGE-2, MAGE-3, MAGE-4, MAGE-5, MAGE-6, MAGE-7, MAGE-8, MAGE-9, MAGE-10, MAGE-11, MAGE-12, MAGE 13, GAGE-1, GAGE-2, GAGE-3, GAGE-4, GAGE-5, GAGE-6, GAGE-7,
10 GAGE-8, BAGE-1, RAGE-1, RAGE-2, RAGE-3, RAGE-4, LB33/MUM-1, DAGE (PRAME), NAG, MAGE-Xp2 (MAGE-B2), MAGE-Xp3, (MAGE-B3), MAGE-Xp4 (MAGE-B4), tyrosinase, brain glycogen phosphorylase, Melan-A, MAGE-C1, MAGE-C2, NY-ESO-1, LAGE-1, SSX-1, SSX-2 (HOM-MEL-40), SSX-4, SSX-5, SCP-1 and CT-7. for example, antigenic peptides characteristic of tumour include
15 those listed in Table A below.

Table A: Exemplary Antigens

Gene	MHC	Peptide	Position	SEQ ID NO:
MAGE-1	HLA-A1	EADPTGHSY	161-169	8
	HLA-Cw16	SAYGEPRKL	230-238	9
MAGE-3	HLA-A1	EVDPIGHLY	168-176	10
	HLA-A2	FLWGPRALV	271-279	11
	HLA-B44	MEVDPIGHLY	167-176	12
BAGE	HLA-Cw16	AARAVFLAL	2-10	13
GAGE-1,2	HLA-Cw16	YRPRPRRY	9-16	14
RAGE	HLA-B7	SPSSNRIRNT	11-20	15
GnT-V	HLA-A2	VLPDVFIRC(V)	2-10/11	16,17
MUM-1	HLA-B44	EEKLIVVLF	exon 2/intron	18
		EEKLSVVLF (wild type)		19

CDK4	HLA-A2	ACDPHSGHFV	23-32	20
		ARDPHSGHFV (wild type)		21
β -catenin	HLA-A24	SYLDSGIHF	29-37	22
		SYLDSGIHS (wild type)		23
Tyrosinase	HLA-A2	MLLAVLYCL	1-9	24
	HLA-A2	YMNGTMSQV	369-377	25
	HLA-A2	YMDGTMSQV	369-377	41
	HLA-A24	AFLPWHRLF	206-214	26
	HLA-B44	SEIWRDIDF	192-200	27
	HLA-B44	YEIWRDIDF	192-200	28
	HLA-DR4	QNILLSNAPLGPQFP	56-70	29
	HLA-DR4	DYSYLQSDPDSFQD	448-462	30
MELAN- AMART-1	HLA-A2	(E)AAGIGILTV	26/27-35	31,32
	HLA-A2	ILTVILGVL	32-40	33
gp100 ^{Pmel 117}	HLA-A2	KTWGQYWQV	154-162	34
	HLA-A2	ITDQVPFSV	209-217	35
	HLA-A2	YLEPGPVTA	280-288	36
	HLA-A2	LLDGTATLRL	457-466	37
	HLA-A2	VLYRYGSFSV	476-485	38
DAGE (PRAME)	HLA-A24	LYVDSLFFL	301-309	39
MAGE-6	HLA-Cw16	KISGGPRISYPL	292-303	40

Other examples will be known to one of ordinary skill in the art (for example, see Coulie, Stem Cells 13:393-403, 1995) and can be used in the invention in a like manner as those disclosed herein. One of ordinary skill in the art can prepare

polypeptides comprising one or more MAGE-A10 peptides and one or more of the foregoing tumour rejection peptides, or nucleic acids encoding such polypeptides, according to standard procedures of molecular biology.

5 Thus polytopes are groups of two or more potentially immunogenic or immune response stimulating peptides which can be joined together in various arrangements (e.g. concatenated, overlapping). The polytope (or nucleic acid encoding the polytope) can be administered in a standard immunization protocol, e.g. to animals, to test the effectiveness of the polytope in stimulating, enhancing and/or provoking
10 an immune response.

The polypeptides can be joined together to directly or via the use of flanking sequences to form polytopes, and the use of polytopes as vaccines is well known in the art (see e.g., Thomson et al. Proc. Acad. Sci USA 92(13):5485-5849), 1995;
15 Gilbert et al, Nature Biotechnol. 15(12):1280:1284, 1997; Thomson et al., J. Immunol. 157(2):822:826, 1996; Tam et al., J. Exp. Med. 171(1): 299-306, 1990). For example, Tam showed that polytopes consisting of both MHC class I and class II binding epitopes successfully generated antibody and protective immunity in a mouse model. Tam also demonstrated that polytopes comprising "strings" of
20 epitopes are processed to yield individual epitopes which are presented by MHC molecules and recognised by CTLs. Thus polytopes containing various numbers and combinations of epitopes can be prepared and tested for recognition by CTLs and for efficacy in increasing an immune response.

25 It is known that tumours express a set of tumour antigens, of which only certain subsets may be expressed in the tumour of any given patient. Polytopes can be prepared which correspond to the different combination of epitopes representing the subset of tumour rejection antigens expressed in a particular patient. Polytopes can be prepared to reflect a broader spectrum of tumour rejection antigens known
30 to be expressed by a tumour type. Polytopes can be introduced to a patient in need of such treatment as polypeptide structures, or via the use of nucleic acid delivery systems known in the art (see. e.g., Allsop et al., Eur. J. Immunol. 26(8):1951-1959, 1996). Adenovirus, pox virus, Ty-virus like particles, adeno-associated virus,

plasmids, bacteria, etc. can be used in such a delivery. One can test the polytope delivery systems in mouse models to determine efficacy of the delivery system. The systems can also be tested in human clinical trials.

5 As part of the immunisation protocols, substances which potentiate the immune response may be administered with the nucleic acid or peptide components of a pharmaceutical composition or a cancer vaccine in accordance with the invention. Such immune response potentiating compound may be classified as either adjuvants or cytokines. Adjuvants may enhance the immunological response by providing a
10 reservoir of antigen (extracellularly or within macrophages), activating macrophages and stimulating specific sets of lymphocytes. Adjuvants of many kinds are well known in the art; specific examples include MPL (SmithKline Beecham), a congener obtained after purification and acid hydrolysis of *Salmonella mimesota* Re 595 lipopolysaccharide. QS21 (SmithKline Beecham), a pure QA-21 saponin purified
15 from *Quillja saponaria* extract, and various water-in-oil emulsions prepared from biodegradable oils such as squalene and/or tocopherol. Cytokines are also useful in vaccination protocols as a result of lymphocyte stimulatory properties. Many cytokines useful for such purposes will be known to one of ordinary skill in the art, including interleukin-12 (IL-12) which have been shown to enhance the protective
20 effects of vaccines (Science 268: 1432-1434, 1995), GM-CSF and IL-18. As envisaged herein, cytokines can be produced *in vivo* by cells transformed or transfected to express nucleic acid molecules coding therefor.

There are a number of additional immune response potentiating compounds that
25 can be used in vaccination protocols. These include co-stimulatory molecules provided in either protein or nucleic acid form. Such co-stimulatory molecules include the B7-1 and B7-2 (CD80 and CD86 respectively) molecules which are expressed on dendritic cells (DC) and interact with the CD28 molecule expressed on the T cell. This interaction provides costimulation (signal 2) to an
30 antigen/MHC/TCR stimulated (signal 1) T cell, increasing cell proliferation and effector function. B7 also interacts with CTLA4 (CD152) on T cells and studies involving CTLA4 and B7 ligands indicate that the B7-CTLA4 interaction can

enhance antitumour immunity and CTL proliferation (Zheng et al., Proc. Nat'l Acad. Sci. USA 95:6284-6289, 1998).

5 B7 typically is not expressed on tumour cells so they are not efficient antigen
presenting cells (APCs) for T cells. Induction of B7 expression would enable the
tumour cells to stimulate more efficiently CTL proliferation and effector function.
A combination of B7/IL-6/IL-12 costimulation has been shown to induce IFN-
gamma and a Th1 cytokine profile in the T cell population leading to further
enhanced T cell activity (Gajewski et al., J. Immunol. 154:5637-5648, 1995). Tumour
10 cell transfection with B7 has been discussed in relation to in vitro CTL expansion
for adoptive transfer immunotherapy by Wang et al., (J. Immunol. 19:1-8, 1986).
Other delivery mechanisms for the B7 molecule would include nucleic acid (naked
DNA) immunization (Kim et al., Nature Biotechnol. 15:7:641-646, 1997) and
recombinant viruses such as adeno and pox (Wendtner et al., Gene Ther. 4:726-
15 735, 1997). These systems are all amenable to the construction and use of
expression cassettes for the coexpression of B7 with other molecules of choice,
such as polypeptides or proteins in accordance with the invention (including
polytopes), or cytokines. These delivery systems can be used for induction of the
appropriate molecules in vitro vaccination situations. The use of anti-CD28
20 antibodies to directly stimulate T cells in vitro and in vivo could also be considered.

Lymphocyte function associated antigen-3 (LFA-3) is expressed on APCs and some
tumour cells and interacts with CD2 expressed on T cells. This interaction induces
T cell IL-2 and IFN-gamma production and can thus complement but not
25 substitute, the B7/CD28 co-stimulatory interaction (Parra et al., J. Immunol.,
158:637-642, 1997; Fenton et al., J. Immunother. 21:95-108, 1998).

Lymphocyte function associated antigen-1 (LFA-1) is expressed on leukocytes and
interacts with ICAM-1 expressed on APCs and some tumour cells. This interaction
30 induces T cell IL-2 and IFN-gamma production and can thus complement but not
substitute, the B7/CD28 co-stimulatory interaction (Fenton et al., 1998). LFA-1 is
thus a further example of a co-stimulatory molecule that could be provided in a
vaccination protocol in the various ways discussed above for B7.

Complete CTL activation and effector function requires Th cell help through the interaction between the Th cell CD40L (CD40 ligand) molecule and the CD40 molecule expressed by DCs (Ridge et al., Nature 393:474, 1998; Bennett et al.,
5 Nature 393:478, 1998; Schoenberger et al., Nature 393:480, 1998). This mechanism of this co-stimulatory signal is likely to involve upregulation of B7 and associated IL-6/IL-12 production by the DC (APC). The CD40-CD40L interaction thus complements the signal 1 (antigen/MHC-TCR) and signal 2 (B7-CD28) interactions.

10 The use of anti-CD40 antibodies to stimulate DC cells directly, would be expected to enhance a response to tumour associated antigens which are normally encountered outside of an inflammatory context or are presented by non-professional APCs (tumour cells). In these situations Th help and B7 costimulation signals are not provided. This mechanism might be used in the context of antigen
15 pulsed DC based therapies or in situations where Th epitopes have not been defined within known tumour associated antigen precursors.

Pharmaceutical compositions in accordance with the present invention can be formulated with conventional pharmaceutically acceptable carriers and excipients,
20 either for systemic or local administration. Such carriers and excipients can be selected without difficulty by those skilled in the art and include those which provide for immediate and sustained release.

The present invention involves the generation of MAGE-specific CTLs from a
25 patient other than MZ2 by MLTC for the first time. A CTL clone (CTL 477A/5) was generated that recognises the nonapeptide (TRA) GLYDGMEHL encoded by *MAGE-A10* in the context of HLA-A2. Its overlapping decapeptide (TRA) GLYDGMEHLI could also sensitise target cells to be lysed by the CTL, but less efficiently. CTL 447A/5 recognised not only autologous tumour cells but *MAGE-*
30 *A10*+ tumour cells from other HLA-A2 patients (Fig. 6), suggesting that GLYDGMEHL is a common TRA presented in tumours expressing *MAGE-A10* and HLA-A2. *MAGE-A10* is expressed in tumours more frequently than previously anticipated. By reverse-transcription-PCR, the expression of *MAGE-A10* gene has

been detected in a variety of tumours, including melanomas, lung cancers, head and neck carcinomas, bladder carcinomas, myelomas, prostatic carcinomas, and (see table 2 below). As observed for other *MAGE* genes, the only normal tissue expressing *MAGE-A10* is testis.

5 Clinical trials have also been under way to treat melanoma patients with peptides derived from *MAGE-A1* and *MAGE-A3*. A few patients showed objective tumour regressions after being immunised with pure peptides, though peptide-specific CTL responses were not detected (Marchand, M., et al., 1995, *Int. J. Cancer*. 63:883- 885).
10 When immunised with peptide-pulsed antigen presenting cells or dendritic cells, quite a few patients developed peptide-specific delayed-type hypersensitivity or CTL responses (Nestle, F.O., et al., 1998, *Nat. Med.* 4:328-332; Mukherji, B., et al., 1995, *Proc. Natl. Acad. Sci. USA*. 92:8078-8082; and Hu, X., et al., 1996, *Cancer Res.* 56:2479-2483). One of the obstacles in cancer immunotherapy is the occurrence of
15 antigen loss tumour variants. Since most tumours expressing *MAGE-A10* also express *MAGE-A1* or/and *MAGE-A3* (F. Brasseur, unpublished data), it is anticipated that addition of peptides in accordance with the present invention in a cocktail vaccination will improve the anti-tumour effect by targeting several different antigens.

20 The following examples show the generation of cytolytic T lymphocytes (CTLs) from patent LB 1751, using MLTC techniques, that lysed specifically autologous tumour cells and produced tumour necrosis factor (TNF) upon stimulation with target cells expressing *MAGE-A10*. The recognition by the CTLs was shown to be
25 restricted by HLA-A2.1 and the antigen was found to be encoded by *MAGE-A10* in the region of nt 547 - 825. From the amino acid sequence corresponding to this region, four peptides were found that had the potential to bind to HLA-A2.1. The expression of *MAGE-A10* has been detected in a variety of tumours, but not in normal tissues except testis and the identified antigenic peptides, therefore, clearly
30 add to the repertoire of antigens that have the potential to be used in anti-tumoural vaccination trials.

Brief description of the Sequences

SEQ. ID. NO. 1 is the amino acid sequence of the protein encoded by the MAGE-A10 gene;

5 SEQ. ID. NO. 2 is the amino acid sequence of the protein encoded for by the MAGE-A8 gene;

SEQ. ID. NO. 3 is the nucleotide sequence of the MAGE-A10 gene;

SEQ. ID. NO. 4 is the nucleic acid sequence of MAGE-A10 cDNA, the region coding for the amino acid sequence in SEQ. ID. NO. 1 lies between a bases 357 and 1466;

10 SEQ. ID. NO. 5 is the nucleotide sequence of the MAGE-A8 gene;

SEQ. ID. No. 6 is a partial sequence of the MAGE-A8 gene as published in WO92/20356, with the codons in the coding portion of the gene identified; and

SEQ. ID. NO. 7 is a partial sequence of the MAGE-A10 gene as published in WO92/20356, with the codons in the coding portion of the sequence identified.

15

Brief description of the Figures

Figure 1. Shows the specific lysis of autologous LB 1751 -MEL cells by CTL 447A/5. Control targets included autologous EBV-transformed lymphoblastoid line LB1751-EBV and NK-sensitive line K562. Chromium release was measured after 4
20 h of incubation of chromium labelled target cells with the CTL at different effector to target ratios.

Figure 2. Shows the HLA-restricted recognition of LB1751-MEL cells by CTL 447A/5. LB1751-MEL cells alone or in the presence of mAbs with the specificities indicated were used to stimulate CTL 447A/5. After 24 h of coculture, production
25 of TNF by the CTL was measured by testing toxicity of the supernatants to TNF-sensitive WEHI-164.13 cells.

Figure 3. Shows the identification of the region coding for the antigenic peptide recognised by CTL 447A/5. PCR fragments of different lengths as indicated were cloned into pcDNAI/Amp and cotransfected into COS-7 cells with gene HLA-A2.1. Transfected cells were incubated for 24 h with CTL 447A/5 and the TNF in
30 the supernatants was measured by its toxicity to WEHI-164.13 cells.

Figure 4. Shows the extent of lysis by CTL 447A/5 of peptide-sensitised LB1751-EBV cells. (A) LB1751-EBV cells pulsed with peptides derived from *MAGE-A10*.

Chromium-labelled autologous EBV-transformed lymphoblastoid cells LB1751-EBV were pulsed for 30 min with peptides as indicated at various concentrations before addition of CTL 447A/5 at an E/T ratio of 20. Chromium release was measured after 4 h. (B) Enhancement by mAb MA2.1 of lysis of LB 1751-EBV cells pulsed with MAGE-A10 peptides. LB1751-EBV cells were pre-treated with or without anti-HLA-A2 antibody MA2.1. The pre-treatment was performed by adding mAb MA2.1 during ⁵¹Cr-labeling. Peptide sensitisation and chromium release assay were carried out as in (A).

Figure 5. Shows the extent of lysis by CTL 447A/5 of LB1751-EBV cells sensitised with peptides derived from MAGE-A8. LB1751-EBV cells were pre-treated with or without anti-HLA-A2 antibody MA2. 1. Ab treatment and peptide sensitisation of the cells and chromium release assay were carried out as in Fig. 4.

Figure 6. Shows the degree of recognition of allogenic tumour cell lines by CTL 447A/5. LB373-MEL (MAGE-A10+), AVL3-MEL (MAGE-A10+) and TT (MAGE-A8+) cell lines derived from HLA-A2 patients were used to stimulate CTL 447A/5. Autologous tumour cell line LB1751-MEL was included as a control. After 24 h of coculture, production of TNF by the CTL was measured by testing toxicity of the supernatants to TNF-sensitive WEHI-164.13 cells.

Example 1

Preparation of CTL Clones against LB1751-MEL and identification HLA-A2.1 as on the presenting MHC molecule.

Melanoma cell line LB1751-MEL was derived from a metastatic melanoma in axillary lymph nodes of a 67-yr-old male patient LB1751 and grown by a method previously described (Van den Eynde, B., et al., 1989, *Int. J. Cancer*. 44:634-640). At passage 4 after the initiation of LB1751-MEL culture, aggregates of typical EBV-transformed lymphoblastoid cells appeared in the supernatant. They were collected and cultured separately to obtain B cell line LB 1751-EBV. Melanoma culture LB1751-MEL was cleared of EBV-transformed B cells by limiting dilution cloning. DNA fingerprint confirmed that LB 1751-MEL and LB 1751-EBV originated from the same patient (data not shown). A panel of CTL clones was generated by MLTC as described previously with minor modifications (Herin, M., et al., 1987, *Int. J.*

Cancer. 39:390-396). Briefly, MLTC was carried out by culturing PBL of patient LB1751 with irradiated LB1751-MEL cells in an 8% CO₂ incubator in Iscove's modified Dulbecco's medium (GIBCO BRL, Gaithersburg, MD) supplemented with 10 mM Hepes buffer, L-arginine (116µg/ml), L-asparagine (36µg/ml), L-glutamine (216g/ml), 10% human serum, and 5 ng/ml of recombinant human IL-7 (rhIL-7) (Genzyme, Cambridge, MA). On day 3, rhIL-2 (Eurocetus, Amsterdam, Netherlands) was added at a final concentration of 25 U/ml. Lymphocytes were restimulated weekly with irradiated LB1751-MEL cells in fresh medium containing 25U/ml of rhIL-2 and 5 ngl/ml of rhIL-7. On day 21, CD8+ T lymphocytes were sorted by using anti-CD8-conjugated MACS magnetic MicroBeads (MACS, Miltenyi Biotec GmbH, Germany) and cloned by limiting dilution. The resulting panel of CTL clones specifically lysed LB1751-MEL cells, but not autologous EBV-transformed B cell line LB 1751-EBV or NK-sensitive cell line K562. Lysis of target cells was tested by chromium release as previously described in (Boon, T., et al., 1980, *J. Exp. Med.* 152:1184-1193) and the results of these tests for representative CTL clone 447A/5 are shown in Fig. 1.

The ability of CTL clone 447A/5 to produce TNF when stimulated with LB1751-MEL cells was confirmed using the technique described in (Traversari, C., et al., 1992, *Immunogenetics*. 35:145-152). Briefly, 2 x 10⁴ tumour cells were grown for 24 h. The medium was discarded and 3,000 CTL were added to the microwells in 100 µl of Iscove's modified Dulbecco's medium supplemented with 10% human serum and 25 U/ml rhIL-2. After 24 h, the supernatant was collected and its TNF content was determined by testing its cytotoxic effect on WEHI 164 clone 13 cells (Espevik, T., et al., 1986, *J. Immunol. Methods*. 95:99-105) in a MTT colorimetric assay (Traversari, C., et al., 1992, *Immunogenetics*. 35:145-152; and Hansen, M. B., et al., 1989, *J. Immunol. Methods*. 119:203-210). Inhibition of TNF production by mAbs W6/32 (anti-HLA class 1) (Bamstable, C.J., et al., 1978, *Cell*. 14:9-20), BB7.2 (anti-HLA-A2) (Parham, P., and F.M. Brodsky, 1981, *Hum. Immunol.* 3:277-299), and B1.23.2 (anti-HLA-B and -C) (Rebai, N., and B. Malissen, 1983, *Tissue Antigens*. 22:107-117) was tested by adding a 1/20 dilution of ascites to the test, and it was found that production of TNF was inhibited by mAbs W6/32 (anti-HLA class I) and BB7.2 (anti-HLA-A2), but not by mAb B1.23.2 (anti-HLA-B, -C) (Fig. 2), indicating that

the target antigen is presented by HLA-A2. The results of the test are set out in Figure 2.

Example 2

5 Identification of the genes encoding the antigen recognised by CTL 447A/5

Because of the high level expression of almost all the *MAGE-A* genes in melanoma cell line LB1751 -MEL (data not shown), the possibility that CTL 447A/5 recognises an antigen encoded by one of the *MAGE-A* genes was tested. COS-7
10 cells were cotransfected with the cDNA of *MAGE-A* genes cloned in expression vector pcDNAI/Amp together with pcDNAI/Amp-A2, a construct encoding the HLA-A2.1. Transfection was performed by the DEAE-dextran-chloroquine method (Seed, B., et al., 1987, *Proc. Natl. Acad. Sci. USA.* 84:3365-3369). Briefly, 2×10^4 COS-7 cells were transfected with 100 ng of plasmid pcDNAI/Amp-A2, a
15 recombinant plasmid containing the HLA-A2.1 gene isolated from a CTL clone of patient SK29 (Wolfel, T., et al., 1993, *Int. J. Cancer.* 55:237-244), and 100 ng of DNA of *MAGE-A* genes cloned in pcDNAI/Amp. The transfectants were grown for 48 hours and then tested for their ability to stimulate TNF production by CTL 447A/5 by the method described in Example 1. The tests revealed that a very significant
20 amount of TNF was produced by CTL 447A/5 when stimulated with COS-7 cells transfected with *MAGE-A10* DNA. Transfectants with *MAGE-A8* cDNA could also stimulate CTL 447A/5 to produce TNF, but less efficiently than those with *MAGE-A10* cDNA. No stimulation was observed with COS-7 cells transfected with HLA-A2.1 alone or with the combination of HLA-A2.1 and any of the other
25 *MAGE-A* genes. The results of these tests are set out in table 1.

Table 1. *Stimulation of CTL 447A/5 by COS-7 cells transfected with HLA-A2.1 and MAGE-A genes*

Stimulator cells	TNF released by CTL 447A/5 (pg/ml)
LB1751-MEL	28
COS	7
COS+HLA-A2.1	4
COS+HLA-A2.1 +	

MAGE-A1	3
MAGE-A2	4
MAGE-A3	4
MAGE-A4	4
MAGE-A6	4
MAGE-A8	30
MAGE-A9	3
MAGE-A10	> 120
MAGE-A11	4
MAGE-A12	2

Control stimulator cells included autologous LB1751-MEL, untransfected COS-7 cells, and COS-7 cells transfected only with HLA-A2.1 gene.

Example 3

5 Identification of the MAGE-A10 Antigenic Peptides.

Fragments of different lengths starting from the initiation codon of *MAGE-10* (nucleotide 1955 in SEQ. ID. NO. 3) were generated by PCR amplification.

The 1.1-kb open reading frame (ORF) of MAGE-A10 was cloned in plasmid vector
10 pcDNA1/Amp (Invitrogen Corporation, Oxon, UK). Three fragments containing the first 270, 546 and 825 nucleotides of the MAGE-A10 open reading frame (ORF) (nucleotides 1955-3064 in SEQ. ID. No. 3) were amplified by PCR using sense primer 5'-GGAATTCATCATGCCTCGAGCTCCAAAGC-3' and three anti-sense primers 5'-GCTCTAGAGCTTAGGCTATCTGAGCACTCTG-3', 5'-
15 GCTCTAGAGCTTAGCACTCGGAGGCTTCACT-3', and 5'-

GCTCTAGAGCTTACCAATCTTGGGTGAGCAG-3' respectively. For PCR amplification *Pfu* DNA polymerase (STRATAGENE, La Jolla, CA) was used. A first denaturation step was done for 5 min at 94°C. The first cycle of amplification was performed for 1 min at 94°C followed by 1 min at 53°C and 1 min at 72°C, and
20 then additional 25 cycles were performed as follows: 1 min at 94°C, 1 min at 65°C, and 1 min at 72°C. Cycling was concluded with a final extension step of 15 min at 72°C.

The PCR products were digested with EcoRI and Xba I, unidirectionally cloned into
25 the EcoRI and Xba 1 sites of plasmid pcDNA1/Amp and transfected into COS-7 cells together with pcDNA1/Amp-A2, using the DEAE-dextran-chloroquine

method described in Example 2. A CTL stimulation assay was carried out with the transfectants in the manner described in Examples 1 and 2. As shown in Fig. 3, the fragment of 825 bp rendered the transfectants capable of stimulating TNF production by CTL 447A/5, and the 546 bp fragment did not, indicating that the sequence coding for the antigenic peptide is located between nt 547 and 825 of the *MAGE-A10* ORF.

In the amino acid sequence corresponding to the nucleotides 547 - 825 there are two nonapeptides, MLLVFGIDV (codons 183-191) and GLYDGMEHL (254-262), which conform to the HLA-A2.1 peptide binding motif, i.e., Leu or Met at position 2 and Leu, Val or Ile at C-terminus (Rammensee, H.G., et al., 1995, *Immunogenetics*. 41:178-228). These two peptides and their overlapping decapeptides were synthesised on solid phase using F-moc for transient NH₂-terminal protection and characterised by mass spectrometry. The peptides were >90% pure, as indicated by analytical HPLC, and used to sensitise autologous lymphoblastoid cell line LB1751-EBV in a chromium release assay as described in (Boon, T., et al., 1980, *J. Exp. Med.* 152:1184-1193) but modified as follows. The target cells were ⁵¹Cr-labeled for 1 h at 37°C and then washed extensively. 1,000 target cells were then incubated in 96-well microplates in the presence of various concentrations of peptides for 30 min at 37°C and CTLs were added at an E/T ratio of 20. Chromium release was measured after 4 h at 37°C.

It was found that the nonapeptide GLYDGMEHL (254-262) and, less efficiently, the decapeptide GLYDGMEHLI (254-263), could sensitise LB1751-EBV cells to lysis by CTL 447A/5 (Fig. 4A). When pre-treated with anti-HLA-A2 antibody MA2.1 for 1 h before peptide sensitisation, LB1751-EBV cells pulsed with both peptides showed a significantly increased sensitivity to lysis by the CTL (Fig. 4B). mAb MA2.1 can facilitate the binding of peptides to HLA-A2 molecules on the cell surface, thereby augmenting lysis of peptide-sensitised target cells by HLA-A2-restricted peptide-specific CTL (Bodmer, H., et al., 1989, *Nature* 342:443-446). Enhancement of peptide binding to the HLA-A2 molecule was achieved by incubation of target cells during ⁵¹Cr-labeling with a 1/5 dilution of hybridoma culture supernatant of mAb MA2.1 (McMichael, A.J., et al., 1980, *Hum. Immunol.* 1:

121-129; and Bodmer, H., et al., 1989, *Nature* 342:443-446). The other two peptides MLLVFGIDV (183-191) and CMLLVFGIDV (182-191) failed to confer recognition by the CTLs, even after LB1751-EBV cells were treated with mAb MA2. 1.

5 **Example 4**

Identification of MAGE-A8 antigen peptides

The sequence of *MAGE-A8*, which is homologous to that of the *MAGE-A10* gene encoding GLYDGMEHL, codes for peptide GLYDGREHS (codons 232-240) that
10 displays two amino acid changes at positions 6 and 9. This peptide and its overlapping decapeptide GLYDGREHSV (232-241) were synthesised by the technique described above. LB1751-EBV cells incubated with either of the peptides, at a concentration of as high as 10 μ M peptide, were not lysed by CTL 447A/5. However, when the peptide concentration was increased to 100 μ M could
15 GLYDGREHS did sensitise LB1751-EBV cells to lysis (Fig. 5). An enhancement of lysis was observed when the LB1751-EBV cells were pre-treated with mAb MA2.1 and pulsed with GLYDGREHS, but not GLYDGREHSV. Enhancement of peptide binding to the HLA-A2 molecule was achieved by incubation of target cells during 51 Cr-labeling with a 1/5 dilution of hybridoma culture supernatant of mAb MA2.1
20 (McMichael, A.J., et al., 1980, *Hum. Immunol.* 1: 121-129; and Bodmer, H., et al., 1989, *Nature* 342:443-446).

Example 5

MAGE-A10+ Allo-tumours Present the Antigen Recognised by CTL 447A15.

25 Using allogenic HLA-A2+ tumour cell lines that express *MAGE-A10* or *MAGEA8* as stimulator cells, a CTL stimulation assay of the type described above was performed to assess the TNF production by CTL 447A/5. Melanoma cell lines LB373-MEL and AVL3-MEL were derived from patients LB373 and AVL, respectively, and cultured in Iscove's modified Dulbecco's medium containing 10%
30 FCS. Medullary thyroid carcinoma cell line TT (ATCC® No.: CRL1803) was obtained from the American Type Culture Collection (Rockville, MD) and maintained in DMEM supplemented with 10% FCS. The results of these assays are set out in Fig. 6 and show that two *MAGE-A10*+ cell lines LB373-MEL and AVL3-

MEL could stimulate CTL 447A/5 to produce TNF, but MAGE-A8+ cell line TT could not. Moreover, AVL3-MEL cells were recognised by CTL 447A/5 less efficiently than LB373-MEL cells, which is consistent with the finding that the transcription level of MAGE-A10 in AVL3-MEL was lower than that in LB373-MEL (Serrano, et, al. manuscript in preparation).

Example 6

MAGE-A10 is Expressed in a Variety of Tumours.

As the expression of MAGE-A10 has been studied only in a small number of tumours, a series of 314 tumours of various histological types were tested by RT-PCR with primers ensuring specificity for gene MAGE-A10. Briefly, reverse-transcription-PCR (RT-PCR) was performed to detect the expression of MAGE-A10 in tumour tissues. Total RNA purification and cDNA synthesis were carried out as previously described (Weynants et al. Int. J. Cancer. 56:826-829, 1994). 1/40th of the cDNA produced from 2µg of total RNA was amplified using sense primer 5'-CACAGAGCAGCACTGAAGGAG-3' and anti-sense primer 5'CTGGGTAAAGACTCACTGTCTGG-3', which yielded a 485-bp specific fragment of MAGE-A10. For PCR, a first denaturation step was done for 4 min at 94° and then 30 cycles of amplification were performed as follows: 1 min at 94°C, 1 min at 65°C, and 1 min at 72°C. Cycling was concluded with a final extension step of 15 min at 72°C. As shown in Table 2, MAGE-A10 was expressed in a number of tumours of various histological types. The expression of some other MAGE genes was also examined by RT-PCR. Of the 71 tumour samples expressing MAGE-A10, all but two expressed simultaneously at least one of genes MAGE-A1, A2, A3, A4 and A6 (data not shown).

Table 2. Expression of *MAGE-A10* in Tumors

Tumor type	Positive samples/ samples tested *
Bladder carcinomas	
Superficial	5/15 (33%)
Infiltrating	5/15 (33%)
Brain tumors	0/9
Breast carcinomas	0/20
Colorectal carcinomas	0/20
Esophageal squamous carcinomas	6/15 (40%)
Head and neck squamous carcinomas	7/20 (35%)
Leukemias	0/25
Lung carcinomas	
Adenocarcinomas	6/15 (40%)
Squamous carcinomas	10/20 (50%)
Melanomas (of cutaneous origin)	
Primary lesions	4/19 (21%)
Metastases	21/45 (47%)
Mesotheliomas	0/4
Myelomas	3/15 (20%)
Neuroblastomas	2/2
Prostatic carcinomas	1/10 (10%)
Renal carcinomas	0/20
Sarcomas	1/15 (7%)
Thyroid carcinomas	0/5
Uterine carcinomas	0/5

* Expression of *MAGE-A10* was tested by RT-PCR on total RNA with specific primers which give a 485-bp product when cDNA is amplified. Percentage of positive samples is shown in parentheses.

SEQ ID NO. 1

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GLTYDGMLSDVQSMPKTGILILILSIIFIEGYCTPEEVIWEALNMMGLYDGMEHLIYGEPRKLLTQDWV
QENYLEYRQVPGSDPARYEFLWGPRAHAEIRKMSLLKFLAKVNGSDPRSFPLWYEEALKDEEERAQDRI
ATTDDTTAMASASSSATGSFSYPE

SEQ ID NO. 2

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LESVIKNYKNHFPDIFSKASECMQVIFGIDVKEVDPAGHSYILVTCLGLSYDGLLGDDQSTPKTGLLII
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SEQ ID NO. 3

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481 acaggcagga agatgaggaa ccctcagggg aatggagtgt tggtgtaaag gggagatatc
541 agccctggac accccacagg gatgacagga tgtggctcct tcttactttt gttttggaat
601 ctcagggagg tgagaacctt gctctcagag ggtgactcaa gtcaacacag ggaacccttc
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SEQ ID NO. 4

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SEQ ID NO. 5

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SEQ ID NO. 6

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SEQ ID NO. 7

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Claims

1. An isolated polypeptide comprising an unbroken sequence of amino acids from SEQ ID. NO. 1, or 2, characterised by an ability to complex with a major histocompatibility complex molecule type HLA-A2, preferably HLA-A2.1.
2. An isolated polypeptide comprising an unbroken sequence of amino acids from SEQ. ID. NO. 1, or 2, characterised by an ability to elicit an immune response from human lymphocytes.
3. A nonapeptide comprising an unbroken sequence of amino acids from SEQ. ID. NO. 1, or 2, wherein the amino acid adjacent to the N-terminal amino acid is L or M, preferably L, and the C-terminal amino acid is L, V, or I, preferably L.
4. A decapeptide comprising a nonapeptide as claimed in claim 3 and, preferably, an unbroken sequence of amino acids from SEQ. ID. NO. 1, or 2.
5. A nonapeptide having the amino acid sequence GLYDGMEHL or GLYDGREHS, preferably GLYDGMEHL.
6. A decapeptide having the amino acid sequence GLYDGMEHLI or GLYDGREHSV, preferably GLYDGMEHLI.
7. An isolated polypeptide of up to about 93 amino acids in length, characterised by comprising a nonapeptide or a decapeptide as claimed in any of claims 3-6.
8. A polypeptide as claimed in claim 7 comprising of an unbroken sequence of amino acids from SEQ. ID. NO. 1, or 2.
9. A polypeptide as claimed in any of the preceding claims, wherein the unbroken sequence is from SEQ. ID. NO. 1.

10. A polypeptide as claimed in any of the preceding claims and capable of eliciting an immune response from human lymphocytes.

5 11. A polypeptide as claimed in claim 10 and capable of eliciting an immune response from human lymphocytes when complexed with a major histocompatibility complex molecule type HLA-A2, preferably HLA-A2.1.

12. A polypeptide as claimed in claim 10 or claim 11, wherein said immune response is an cytolytic response from human T-lymphocytes.

10

13. An isolated polypeptide or protein comprising a polypeptide as claimed in any of claims 1-12, wherein the amino acid sequence of said isolated polypeptide or protein is not that set out in either of SEQ. ID. NOs. 1 and 2 or that coded for by nucleotides 334-918 of SEQ. ID. NO. 7.

15

14. An isolated polypeptide or protein which is a functionally equivalent homologue to a polypeptide or protein as claimed in any of claims 1-13, wherein the amino acid sequence of said isolated polypeptide or protein is not that set out in either of SEQ. ID. NOs. 1 and 2 or that coded for by nucleotides 334-918 of SEQ. ID. NO. 7.

20

15. An isolated nucleic acid molecule comprising a nucleotide sequence coding for a polypeptide or protein as claimed in any of claims 1-13, or a complimentary nucleotide sequence, wherein said nucleotide sequence is not that set out in any of SEQ. ID. NOs. 3, 4, 5, 6 or 7.

25

16. A nucleic acid molecule as claimed in claim 15 and comprising an unbroken sequence of nucleotides from SEQ. ID. NO. 3, 4 or 5, or a complimentary sequence, or an RNA transcript of said nucleic acid molecule.

30

17. A nucleic acid molecule as claimed in claim 15 or claim 16, wherein said nucleotide sequence encodes a plurality of epitopes or a polytope.

18. An expression vector comprising a nucleic acid molecule as claimed in any of claims 15-17 operably linked to a promoter.
19. An expression vector as claimed in claim 18, further comprising a nucleotide
5 sequence coding for a major histocompatibility complex molecule type HLA-A2,
preferably HLA-A2.1, a cytokine or a co-stimulatory molecule, or a bacterial or viral
genome or a portion thereof.
20. A host cell transformed or transfected with an expression vector as claimed
10 in claim 18 or claim 19.
21. A host cell as claimed in claim 20, transformed or transfected with an
expression vector coding for a major histocompatibility complex molecule type
HLA-A2, preferably HLA-A2.1, a cytokine or a co-stimulatory molecule.
22. A polypeptide-binding agent which selectively binds or is specific for an
15 isolated polypeptide or protein as claimed in any of claims 1-14.
23. A polypeptide-binding agent as claimed in claim 22, comprising an antibody,
20 preferably a monoclonal antibody or an antibody fragment specific for an isolated
polypeptide as claimed in any of claims 1-14.
24. A polypeptide-binding agent as claimed in claim 22 or claim 23 which
selectively binds or is specific for a complex of a polypeptide as claimed in any of
25 claims 1-14 and a major histocompatibility complex molecule type HLA-A2,
preferably HLA-A2.1, but which does not bind said major histocompatibility
molecule alone.
25. A polypeptide-binding agent as claimed in any of claims 22-24, comprising a
30 cytolytic T-cell which is specific for a complex of a polypeptide as claimed in any of
claims 1-14 and a major histocompatibility complex molecule type HLA-A2,
preferably HLA-A2.1.

26. A polypeptide or protein as claimed in any of claims 1-14, an isolated nucleic acid molecule as claimed in any of claims 15-17, an expression vector as claimed in either of claims 18 or 19, a host cell as claimed in either of claims 20 or 21, or a polypeptide binding agent as claimed in any of claims 22-25, for use in the therapy, prophylaxis or diagnosis of tumours.

27. A pharmaceutical composition for the prophylaxis, therapy or diagnosis of tumours comprising a polypeptide or protein as claimed in any of claims 1-14, a nucleic acid molecule as claimed in any of claims 15-17, an expression vector as claimed in either of claims 18 or 19, a host cell as claimed in either of claims 20 or 21, or a polypeptide binding agent as claimed in any of claims 22-25, optionally in admixture with a pharmaceutically acceptable carrier and optionally further comprising a major histocompatibility molecule type HLA-A2, preferably HLA-A2.1.

15

28. A pharmaceutical composition for the prophylaxis, therapy or diagnosis of tumours comprising a polypeptide or protein as claimed in any of claims 1-14 complexed with a major histocompatibility molecule, HLA, and presented on the surface of an APC, preferably a dendritic cell, wherein said complex is formed by pulsing said APC with polypeptide or protein.

20

29. A cell, preferably an APC, and more preferably, a dendritic cell, which has been pulsed with a polypeptide or protein as claimed in any of claims 1-14 to present on its surface said polypeptide or protein as a complex with a major histocompatibility molecule, HLA.

25

30. A pharmaceutical composition as claimed in any of claims 27 and 28 further comprising a co-stimulatory molecule.

31. A method of diagnosing disease, preferably cancer, comprising contacting a biological sample isolated from a subject with an agent that is specific for a polypeptide or protein as claimed in any of claims 1-14, or a nucleic acid molecule as claimed in any of claims 15-17, and assaying for interaction between the agent

30

and any of the polypeptide, protein or nucleic acid molecule either free in or forming an integral part of the sample as a determination of the disease.

32. A method as claimed in claim 31, wherein the agent is a polypeptide-binding
5 agent as claimed in any of claims 22-25.

33. A method of producing a cytolytic T-cell culture reactive against tumour cells, comprising removing a lymphocyte sample from an individual and culturing the lymphocyte sample with a polypeptide or protein as claimed in any of claims 1-
10 14, an expression vector as claimed in either of claims 18 or 19, or a host cell as claimed in either of claims 20 or 21.

34. A product comprising cytolytic T-cells reactive against a tumour cell expressing an antigen comprising a polypeptide or protein as claimed in any of
15 claims 1 to 14, for use in the prophylaxis, therapy or diagnosis of tumours.

35. A product as claimed in claim 34 and obtained or obtainable by a method as claimed in claim 33.

20 36. A method of treating tumours in a patient comprising administering a composition as claimed in any of claims 26, 27, 28, 30, 34 or 35 to the patient in an amount effective to control or prevent tumour growth .

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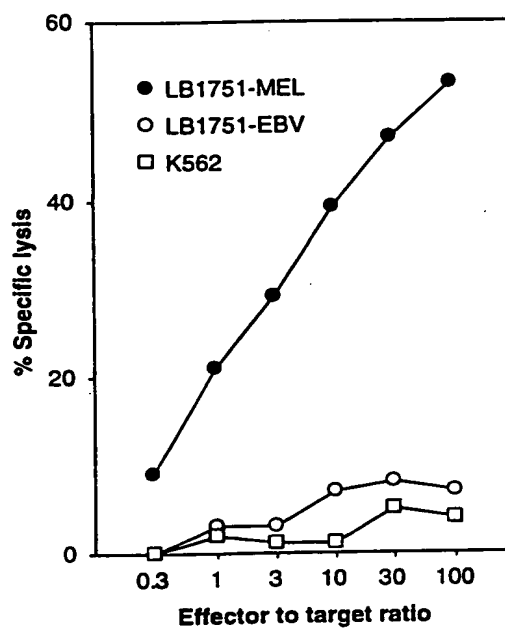


Fig. 1

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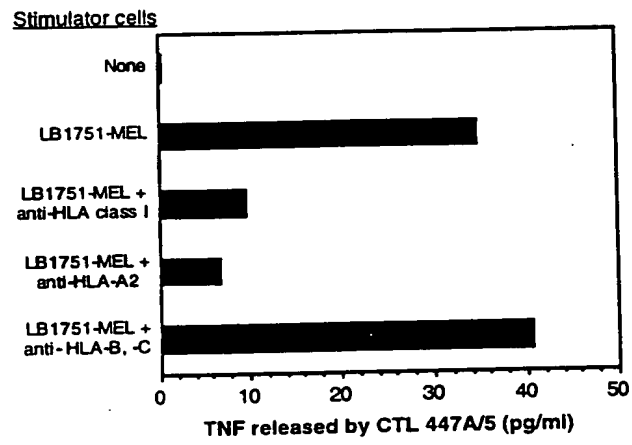


Fig. 2

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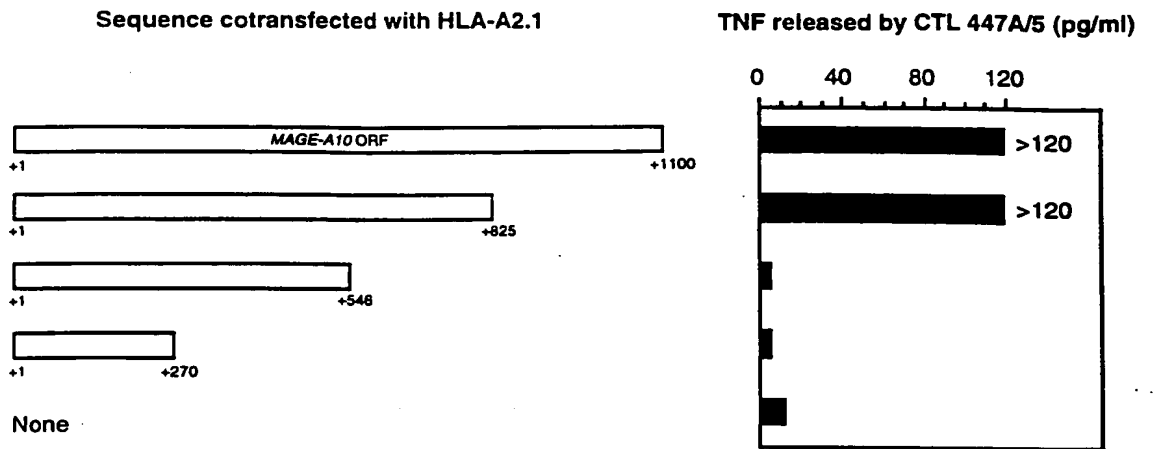


Fig. 3

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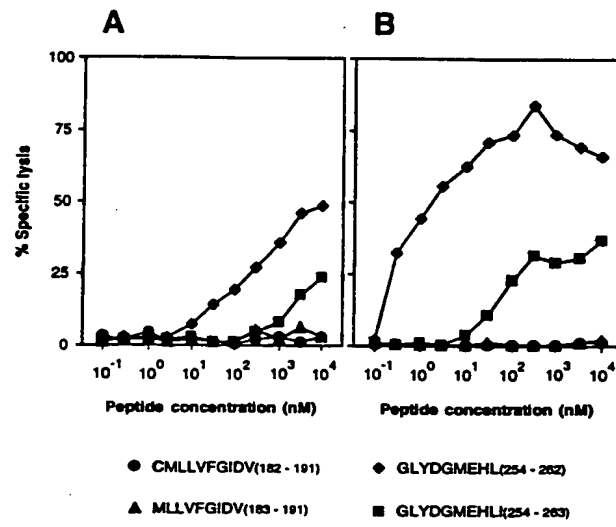


Fig. 4

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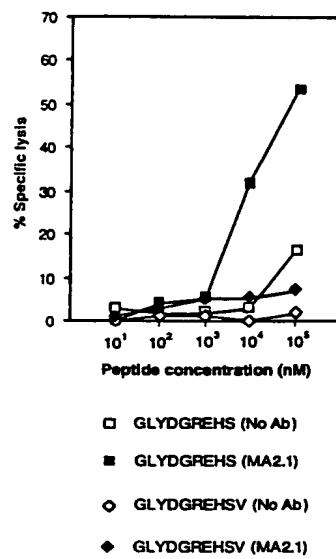


Fig. 5

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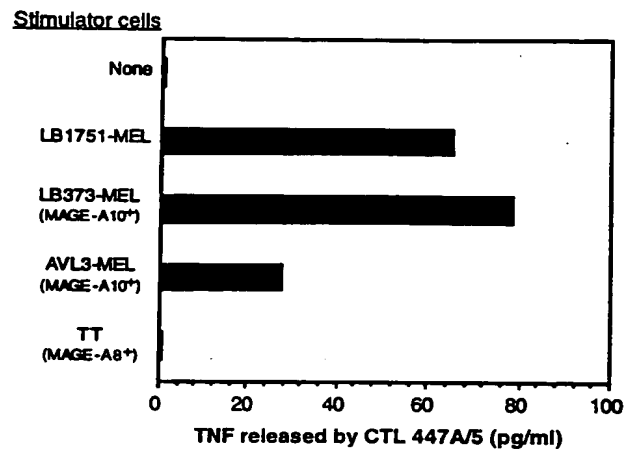


Fig. 6

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